Bioorganic & Medicinal Chemistry Letters

Bioorganic & Medicinal Chemistry Letters 16 (2006) 834-838

Androstenediol analogs as ER-β-selective SERMs

Timothy A. Blizzard,^{a,*} Candido Gude,^a Jerry D. Morgan, II,^a Wanda Chan,^a Elizabeth T. Birzin,^a Marina Mojena,^c Consuelo Tudela,^c Fang Chen,^b Kristin Knecht,^b Qin Su,^b Bryan Kraker,^b Ralph T. Mosley,^b Mark A. Holmes,^a Nandini Sharma,^a Paula M. D. Fitzgerald,^a Susan P. Rohrer^a and Milton L. Hammond^a

^aMerck Research Laboratories, PO Box 2000, Rahway NJ 07065, USA
^bMerck Research Laboratories, Sunnytown Pike, West Point, PA 19486, USA
^cMerck Research Laboratories-CIBE, Madrid, Spain

Received 4 October 2005; revised 4 November 2005; accepted 4 November 2005 Available online 23 November 2005

Abstract—A series of 19-substituted androstenediol derivatives was prepared. Some of the novel analogs were surprisingly potent and selective ligands for ER-β.
© 2005 Elsevier Ltd. All rights reserved.

The clinical significance of the selective estrogen receptor modulators (SERMs)¹ generated substantial interest in the discovery of new SERMs.² Reports of a second estrogen receptor subtype³ prompted interest in the development of receptor subtype-selective SERMs, and both ER- α^4 and ER- β^5 selective SERMs have since been described in the literature. Although these efforts have focused on non-steroid structures, several steroidal SERMs have also been reported.⁶ We have previously reported the discovery of spiroindenes (e.g., 1)⁷ as non-subtype-selective SERMs and of dihydrobenzoxathiins (e.g., 2) as ER- α selective SERMs.^{5b,c,d}

HO 1 2

Naturally, we were also interested in discovering ER-β-selective ligands. High-throughput screening^{8a} of the Merck sample collection resulted in the discovery of 19-methylene-androstenediol (3) as a potent and highly

Keywords: SERMs; SERAMs; Estrogen; Androstenediol.

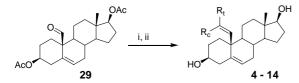
selective ligand for ER-β. 8b An old patent 6d describes 3 as having anabolic-androgenic and anti-estrogen activities (and other activities) but provides no binding data and does not discuss estrogen receptor subtype selectivity.

The excellent binding affinity and ER-β selectivity of 3 (Table 1) prompted us to initiate a medicinal chemistry program to explore the SAR of this remarkable lead. We decided to focus initially on substitution of the 19,20 olefin. A series of olefin-substituted analogs was prepared and evaluated as summarized below (Schemes 1-6).9 Wittig olefination of aldehyde **29**¹⁰ followed by deacylation afforded the C-20-substituted olefin analogs 4-14 in low to moderate yields (Scheme 1). As expected, the Wittig reaction afforded a mixture of olefin isomers from which the cis and trans isomers of the product could be isolated by careful chromatography. Interestingly, the outcome of Wittig olefination was dependent on the nature of the protecting groups at C-3 and C-17. Substitution of TBDMS or THP for the acetate-protecting groups of 29 resulted in significant changes in yield and cis:trans product ratio in many cases. Despite numerous attempts, we were unable to prepare analogs

^{*}Corresponding author. Tel.: +1 732 594 6212; fax: +1 732 594 9556; e-mail: tim_blizzard@merck.com

Table 1. Biodata

Compound	R	R ¹⁷	ER binding (IC ₅₀ , nM) ¹²			ER transactivation (EC ₅₀ , nM) ¹³			AR (IC ₅₀ , nM) ¹⁴	
			hERα	hERβ	α/β	α	β	α/β	AR	AR/ERβ
3	CH=CH ₂	Н	2236	11	212	982	4.1	246	33	3
4	cis-CH=CHF	Н	4280	25	171	622	3.8	164	11	0.4
5	trans-CH=CHF	Н	4165	22	188	324	8.7	37	85	4
6	cis-CH=CHCl	Н	6690	83	80	>1000	91	>11	172	2
7	trans-CH=CHCl	Н	5600	122	46	603	68	9	1436	12
8	cis-CH=CHBr	Н	>10,000	212	47	>1000	96	>10	20	0.1
9	trans-CH=CHBr	Н	3984	210	19	>1000	67	15	120	0.6
10	cis-CH=CHOMe	Н	3349	303	11	3237	188	17	1402	5
11	trans-CH=CHOMe	Н	>10,000	1299	8	583	827	0.7	4617	4
12	cis-CH=CHMe	Н	1956	16	126	344	21	16	718	45
13	trans-CH=CHMe	Н	1164	96	12	47	39	1	843	9
14	cis-CH=CHEt	Н	3042	361	8	719	252	3	3767	10
15	CH ₂ CHO	Н	>10,00	1619	>6	>1000	>1000	1	>5000	_
16	$CH_2C=CH_2$	Н	1738	128	14	>1000	148	7	1768	12
17	CH_2CH_3	Н	2329	16	144	_	_	_	88	6
18	CH ₂ CH ₂ CH ₃	Н	5155	132	39	_	_	_	1769	14
19	C≡CH	Н	425	17	25	160	8	20	1761	104
20	$C \equiv CCH_3$	Н	8419	381	22	1000	171	6	3733	10
21	$CH_2C \equiv CH$	Н	4643	286	16	984	153	6	1717	6
22	CH_3	Н	210	10	21	26	6	4	212	21
23	CH ₂ OH	H	6324	1754	4	>1000	641	>1	>5000	>2.9
24	CHO	Н	>10,000	230	>43	844	78	11	>5000	>22
25	$CH=CH_2$	Me	1053	25	42	494	31	16	169	7
26	$CH=CH_2$	C≡CH	147	14	10	129	24	5	287	20
27	CH_3	H	>10,000	>10,000	1	_	_	_	2.7	< 0.0002
28	$CH=CH_2$	H	>10,000	782	>12	>1000	379	>2.6	7.9	0.01
17β-estradiol	_	_	1.4	1.2	1.2	0.75	2.1	2.8	19.1	16



Scheme 1. Reagents: (i) RtRcC=PPh3; (ii) MeOH, KOH.

Scheme 2. Reagents: (i) $\rm H_2$ (50 psi), 5% Rh/C, EtOAc, EtOH; (ii) TBAF, THF.

of 3 with substitution at C-19. For example, attempts to prepare the 19-methyl-substituted analog of 3 by olefination of the methyl ketone analog of aldehyde 29 failed due to the complete unreactivity of the ketone to both the Wittig and Tebbe methylenation reagents, presumably due to steric hindrance at C-19. Alternative approaches were also unsuccessful.

Scheme 3. Reagents: (i) Ph₃PCH₂Cl Cl, *n*BuLi; (ii) KOH, MeOH, (iii) DHP, pyridinium tosylate, 90% from **29**; (iv) *n*BuLi, 56%; (v) pyridinium tosylate, MeOH, 78%.

To determine whether a double bond at C-19,20 was the optimum degree of unsaturation, we prepared the corresponding saturated analog 17 and the acetylene 19. For comparison, we also evaluated the parent compound androstenediol (22). The ethyl analog 17 was prepared by hydrogenation of 30 (Scheme 2). This poorly reproducible reaction proved to be somewhat more difficult than anticipated, probably due to the hindered nature of the C-19,20 olefin.

Scheme 4. Reagents: (i) MeOC=PPh₃; (ii) MeOH, pyridinium tosylate; (iii) HCl; (iv) DHP, pyridinium tosylate; (v) H₂C=PPh₃; (vi) MeOH, pyridinium tosylate; (vii) H₂ (50 psi), 5% Rh/C, EtOAc, EtOH.

Scheme 5. Reagents: (i) TBDMS-Cl, imidazole, DMF, 60%; (ii) TPAP, NMO, CH₂Cl₂; (iii) R¹⁷MgCl (R¹⁷=Me) or R¹⁷Li (R¹⁷=CCH), THF; (iv) TBAF, THF.

Scheme 6. Reagents: (i) NaOH, MeOH; (ii) H₂CrO₄; (iii) NaOH, MeOH.

The synthesis of acetylene 19 began with conversion of aldehyde 29 to a mixture of cis and trans chlorides 31. Reaction of 31 with *n*-butyllithium and deprotection afforded acetylene 19 in modest overall yield (Scheme 3). Modification of this route (reaction of the acetylene anion intermediate with iodomethane) afforded the methyl acetylene 20.

The extended analog 16 was prepared by olefination of extended aldehyde 15 which, in turn, was prepared by olefination of 33, the THP-protected analog of 29. Hydrogenation of 16 afforded the propyl analog 18 (Scheme 4). The extended acetylene 21 was prepared by subjecting the protected analog of extended aldehyde 15 to the sequence outlined in Scheme 3 for the synthesis of acetylene 19.

We also prepared the 17-methyl (25) and 17-ethynyl (26) analogs of 3 by reaction of ketone 32 (prepared from 3

by selective silylation of the 3-hydroxyl followed by oxidation of the 17-hydroxyl to the ketone) with either MeMgBr or lithium TMS-acetylide followed by deprotection to afford **25** or **26** (Scheme 5).

The 3-keto-delta-4 analog 28 was prepared by selective deacetylation of 34, an intermediate in the preparation of 3, followed by oxidation of the C-3 hydroxyl and deprotection (Scheme 6).

The compounds were evaluated first in an ER ligand binding assay (Table 1).¹² All were ER-β selective although the degree of selectivity varied considerably. The olefin-substituted analogs 4-14 were generally less effective than 3 as ER-β ligands although the fluoro analogs 4 and 5, and the cis-methyl analog 12 were comparable to 3 in selectivity and binding affinity. cis-substitution at C-20 was clearly preferred and smaller substituents were better than larger ones. Interestingly, substitution at C-17 (compounds 25 and 26) had little effect on ER-β binding but substantially improved binding to ER- α (especially with 17-ethynyl substitution) resulting in lower ER-β selectivity. Both the saturated derivative 17 and the acetylene analog 19 retained the ER- β binding affinity of 3. However, the acetylene was a surprisingly better ligand for ER α so 19 was less selective for ER- β than either 3 or 17. The extended olefin was clearly less effective than 3 and the same trend was also observed in the saturated (18 vs 17) and acetylene (20 and 21 vs 19) cases. Androstenediol (22) was an excellent ligand for ER-β but was also a good ligand for ER- α and so was much less selective than 3. The oxygenated analogs 23 and 24 were much weaker ligands.

The compounds were also evaluated in a cell-based transactivation assay. With the exception of the *cis*-fluoro analog **4**, the olefin-substituted analogs **4–14** were somewhat less active than **3** in this assay and were less selective as well. Once again, *cis*-substitution at C-20 was preferred. In general, ER- β selectivity was lower in this assay compared to the binding assay. The 17-substituted analogs **25** and **26** had good ER- β potency but were less selective than **3**.

One of our concerns when we initiated this project was the possibility of off-target activity mediated by binding to other steroid receptors, particularly the androgen receptor (AR). The lead compound 3 was a 33 nM ligand for AR but had no significant binding $(IC_{50} > 3000 \text{ nM})$ to either the glucocorticoid receptor (GR) or the progesterone receptor (PR). New compounds were therefore routinely evaluated in an AR binding assay. 14 The 20-cis-methyl analog 12 had the best AR/ERB binding ratio of the olefin-substituted analogs 4-14, and was perhaps the most interesting compound overall, with a 15-fold improvement in selectivity for ERβ over AR (relative to 3) and only a minimal loss in selectivity for ER β over ER α . Overall, the acetylene 19 had the best AR/ERβ selectivity. However, selectivity for ER- β over ER- α was somewhat less for this compound. As expected, testosterone (27) and the testosterone analog 28 were excellent ligands for the androgen receptor.

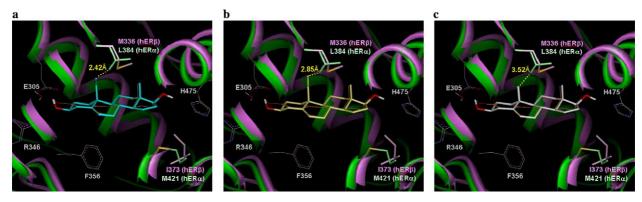


Figure 1. Superposition of estradiol (orange) in the context of hER- α (green) with (a) olefin 3 (cyan) (manually docked into a homology model of ER- β ; proposed binding mode for 3 was later confirmed crystallographically); (b) acetylene analog 19 (yellow); and (c) androstenediol 22 (white) in the context of hER- β (purple) complexed with compound 3. Unless otherwise indicated, residue numbering is that of hER- β . See Ref. 15 for modeling details.

Molecular modeling¹⁵ of compounds 3, 17, 19, and 22 provides a rationale for the ER-\beta selectivity observed with 3 and 17 (Fig. 1). The terminal carbon of the vinyl group for 3 is approximately, 2.4 Å away from Leu384 in ER- α , which is about 1.0 A closer than the distance to the corresponding Met336 in ER-β. Similarly, for the saturated analog 17, the terminal carbon of the ethyl group is about 2,3 Å away from Leu384 in ER-α, which is about 1.0 Å closer than the distance to the corresponding Met336 in ER-β (not shown in Fig. 1). The proposed negative steric interaction with Leu384 in hER- α is consistent with the SAR for this series of compounds for which ER-B selectivity is gained through a decrease in ER-α affinity. Compounds 19 and 22, by comparison, have smaller substituents which are further removed from Leu384 (2.9 and 3.5 Å, respectively) and would be expected to have less of a negative interaction with ER-α. Both lose ER-β selectivity relative to 3 and 17 through an increase in ER- α affinity.

In conclusion, the androstenediol analogs reported herein exhibit a range of binding affinities and selectivities for the estrogen receptors (up to ~ 200 -fold selective for ER- β over ER- α). Additional studies will be reported in future communications.

Acknowledgments

The authors thank Susanne Miranda and Joe Laquidara for the large-scale preparation of **29**.

References and notes

- (a) Jordan, V. C. J. Med Chem. 2003, 46, 883; (b) Lonard,
 D. M.; Smith, C. L. Steroids 2002, 67, 15.
- (a) Jordan, V. C. J. Med. Chem. 2003, 46, 1081; (b) Veeneman, G. H. Curr. Med. Chem. 2005, 12, 1077.
- (a) Kuiper, G. G. J. M.; Enmark, E.; Pelto-Kuikko, M.; Nilsson, S.; Gustafsson, J. A. *Proc. Natl. Acad. Sci. U.S.A.* 1996, 93, 5925; (b) Mosselman, S.; Polman, J.; Dijkema, R. *FEBS Lett.* 1996, 392, 1996.

- 4. (a) Mortensen, D. J.; Rodriguez, A. L.; Carlson, K. E.; Sun, J.; Katzenellenbogen, B. S.; Katzenellenbogen, J. A. J. Med. Chem. 2001, 44, 3838; (b) Kim, S.; Wu, J. Y.; Birzin, E. T.; Frisch, K.; Chan, W.; Pai, L.; Yang, Y. T.; Mosley, R. T.; Fitzgerald, P. M. D.; Sharma, N.; DiNinno, F.; Rohrer, S.; Schaeffer, J. M.; Hammond, M. L. J. Med. Chem. 2004, 47, 2171; (c) Blizzard, T. A.; DiNinno, F.; Morgan, J. D., II; Chen, H. Y.; Wu, J. Y.; Kim, S.; Chan, W.; Birzin, E. T.; Yang, Y.; Pai, L.; Fitzgerald, P. M. D.; Sharma, N.; Li, Y.; Zhang, Z.; Hayes, E. C.; DaSilva, C. A.; Tang, W.; Rohrer, S. P.; Schaeffer, J. M.; Hammond, M. L. Bioorg. Med. Chem. Lett. 2005, 15, 107; (d) Blizzard, T. A.; DiNinno, F.; Chen, H. Y.; Kim, S.; Wu, J. Y.; Chan, W.; Birzin, E. T.; Yang, Y.; Pai, L.; Hayes, E. C.; DaSilva, C. A.; Rohrer, S. P.; Schaeffer, J. M.; Hammond, M. L. Bioorg. Med. Chem. Lett. 2005, 15, 3912.
- 5. (a) Henke, B. R.; Consler, T. G.; Go, N.; Hale, R. L.; Hohman, D. R.; Jones, S. A.; Lu, A. T.; Moore, L. B.; Moore, J. T.; Orband-Miller, L. A.; Robinett, R. G.; Shearin, J.; Spearing, P. K.; Stewart, E. L.; Turnbull, P. S.; Weaver, S. L.; Williams, S. P.; Wisely, G. B.; Lambert, M. H. J. Med. Chem. 2002, 45, 5492; (b) Schopfer, U.; Schoefter, P.; Bischoff, S. F.; Nozulak, J.; Feuerbach, D.; Floersheim, P. J. Med. Chem. 2002, 45, 1399; (c) De Angelis, M.; Stossi, F.; Carlson, K. A.; Katzenellenbogen, B. S.; Katzenellenbogen, J. A. J. Med. Chem. 2005, 48, 1132; (d) Malamas, M. S.; Manas, E. S.; McDevitt, R. E.; Gunawan, I.; Xu, Z. B.; Collini, M. D.; Miller, C. P.; Dinh, T.; Henderson, R. A.; Keith, J. C.; Harris, H. A. J. Med. Chem. 2004, 47, 5021; (e) Manas, E. S.; Unwalla, R. J.; Xu, Z. B.; Malamas, M. S.; Miller, C. P.; Harris, H. A.; Hsiao, C.; Akopian, T.; Hum, W. T.; Malakian, K.; Wolfrom, S.; Bapat, A.; Bhat, R. A.; Stahl, M. L.; Somers, W. S.; Alvarez, J. C. J. Am. Chem. Soc. 2004, 126, 15106; (f) Vu, A. T.; Cohn, S. T.; Manas, E. S.; Harris, H. A.; Mewshaw, R. E. Bioorg. Med. Chem. Lett. 2005, 15; (g) Yang, W.; Wang, Y.; Ma, Z.; Golla, R.; Stouch, T.; Seethala, R.; Johnson, S.; Zhou, R.; Gungor, T.; Feyen, J. H. M.; Dickson, J. K. Bioorg. Med. Chem. Lett. 2004, 14, 2327; (h) De Angelis, M.; Stossi, F.; Waibel, M.; Katzenellenbogen, B. S.; Katzenellenbogen, J. A. Bioorg. Med. Chem. Lett. 2005, 15, 6529; (i) Mewshaw, R. E.; Edsall, R. J.; Yang, C.; Manas, E. S.; Xu, Z. B.; Henderson, R. A.; Keith, J. C.; Harris, H. A. *J. Med. Chem.* **2005**, *48*, 3953.
- (a) Hegele-Hartung, C.; Siebel, P.; Peters, O.; Kosemund,
 D.; Muller, B.; Hillisch, A.; Walter, A.; Kraetzschmar, J.;
 Fritzemeier, K. H. Proc. Natl. Acad. Sci. U.S.A. 2004, 101,

- 5129; (b) Hanson, R. N.; Friel, C. J.; Dilis, R.; Hughes, A.; DeSombre, E. R. *J. Med. Chem.* **2005**, *48*, 4300; (c) Zhang, J.; Labaree, D. C.; Hochberg, R. B. *J. Med. Chem.* **2005**, *48*, 1428(d) Bowers, A. US 3,102,127, 1963.
- (a) Blizzard, T. A.; Morgan, J. D.; Mosley, R. T.; Frisch, K.; Birzin, E. T.; Rohrer, S. P.; Hammond, M. L. Bioorg. Med. Chem. Lett. 2003, 13, 479.
- (a) Peekhaus, N. T.; Ferrer, M.; Chang, T.; Kornienko, O.; Schneeweis, J. E.; Smith, T. S.; Hoffman, I.; Mitnaul, L. J.; Chin, J.; Fischer, P. A.,; Blizzard, T. A.; Birzin, E. T.; Chan, W.; Inglese, J.; Strulovici, B.; Rohrer, S. P.; Schaeffer, J. M. Assay Drug Dev. Technol. 2003, 1, 789(b)
 (b) Birzin et al. manuscript in preparation.
- 9. All new compounds were characterized by LC-MS and 400, 500, or 600 MHz ¹H NMR.
- Fajkos, J.; Pouzar, V.; Veres, K. Coll. Czech. Chem. Commun. 1990, 55, 2086.
- 11. Androstenediol is commercially available.
- 12. The IC $_{50}$ values were generated in an estrogen receptor ligand binding assay. This scintillation proximity assay was conducted in NEN Basic Flashplates using tritiated estradiol and full length recombinant human ER α and ER β proteins. Compounds were evaluated in duplicate in a single assay. In our experience, this assay provides IC $_{50}$ values that are reproducible to within a factor of 2–3.

- 13. This assay was run at MRL-CIBE in Spain using the procedure described by Barkhem et al: Barkhem, T.; Carlsson, B.; Nilsson, Y.; Enmark, E.; Gustafsson, J.; Nilsson, S. *Mol. Pharmacol.* **1998**, *54*, 105.
- Chen, F.; Knecht, K.; Leu, C.; Rutledge, S. J.; Scafonas, A.; Gambone, C.; Vogel, R.; Zhang, H.; Kasparcova, V.; Bai, C.; Harada, S.; Schmidt, A.; Reszka, A.; Freedman, L. J. Steroid Biochem. Mol. Biol. 2004, 91, 247.
- 15. (a) Models of 3 using crystallographically determined estradiol cocrystallized with hERa(pdb:1ERE) were built and docked into a homology model of hERβ. The proposed binding mode was later confirmed crystallographically (Fitzgerald et al., manuscript in preparation). Models 17, 19, and 22 were based on the crystallographic coordinates of 3 as cocrystallized with hERβ. Energy minimization for all of the models within the context of the hERB receptor (3 cocrystallized) was accomplished by rigidly fixing all residues, except for side chains which fell within 5 Å of the modeled ligand which were allowed to minimize in conjunction with the ligand. All minimizations were conducted using the MMFFs forcefield^{15b} with a distance dependent dielectric model of 2r; (b) Halgren, T. A. J. Comp. Chem. **1999**, 20, 730.